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# Identification of candidate genes for drought stress tolerance in rice by the integration of a genetic (QTL) map with the rice genome physical map\*

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**Abstract:** Genetic improvement for drought stress tolerance in rice involves the quantitative nature of the trait, which reflects the additive effects of several genetic loci throughout the genome. Yield components and related traits under stressed and well-water conditions were assayed in mapping populations derived from crosses of Azucena×IR64 and Azucena×Bala. To find the candidate rice genes underlying Quantitative Trait Loci (QTL) in these populations, we conducted in silico analysis of a candidate region flanked by the genetic markers RM212 and RM319 on chromosome 1, proximal to the semi-dwarf (sd1) locus. A total of 175 annotated genes were identified from this region. These included 48 genes annotated by functional homology to known genes, 23 pseudogenes, 24 *ab initio* predicted genes supported by an alignment match to an EST (Expressed sequence tag) of unknown function, and 80 hypothetical genes predicted solely by *ab initio* means. Among these, 16 candidate genes could potentially be involved in drought stress response.

#### INTRODUCTION

Drought stress is a major constraint to rice (*Oryza sativa*) production and yield stability in rained ecosystems (Dey and Upadhyaya, 1996). Rice must be made more drought tolerant, but this is a somewhat contradictory objective considering that rice is most commonly grown under flooded conditions. Achieving drought tolerance in rice will require a deeper understanding of the possible physiological mechanisms available for water stress tolerance and the identification of favorable alleles for introgression

into rice varieties that otherwise suit specific environments. Quantitative Trait Loci (QTL) are useful starting points for identifying such alleles. QTL relating to drought tolerance have been identified in rice, barley, and grain sorghum (Price *et al.*, 2002a).

However, positionally cloning a QTL gene locus normally requires fine scale mapping with large mapping populations across many seasons. Availability of the whole rice genome sequences (Goff *et al.*, 2002; Yu *et al.*, 2002; Sasaki *et al.*, 2002; Feng *et al.*, 2002) provides a new tool for this task, along with a means of characterizing their associated molecular functions. In this study, we exploited this new source of data by anchoring drought stress tolerance QTL maps reported by other researchers (Lafitte *et al.*, 2002; Price *et al.*, 2002a; 2002b; 2002c; Causse *et al.*,

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1994; Harushima *et al.*, 1998) to a rice physical and sequence map using sequence based markers. We then identified genes as candidates with respect to position.

Drought stress most severely impacts yield when applied during the reproductive stage of the rice plant. The sd1 is one of the most important genes in rice, whose recessive character results in a shortened culm with improved lodging resistance and a harvest index (Jennings, 1964). The rice height-related QTL was shown to be tightly linked to restricted fragment length polymorphism (RFLP) markers RZ730 and RG810 on the long arm of chromosome 1 (Li et al., 2003), consisting of the QTL region of drought resistance (Lafitte et al., 2002). As further evidence for drought tolerance gene candidacy, we aligned EST sequences from an available drought stressed panicle library onto our candidate gene structures. We also assessed gene candidacy based on literature support.

#### MATERIALS AND METHODS

### Genetic map for IR64×Azucena and Bala× Azucena QTL mapping populations

Published genetic mapping data from a double haploid mapping population, IR64×Azucena (Temnykh et al., 2001) and Bala×Azucena (Price et al., 2002b) were used as starting points for this study. IR64×Azucena map included 93 amplified fragment length polymorphism (AFLP), 481 simple sequence repeat (SSR), 15 'known' genes, 1 isozyme, a restriction fragment length polymorphism (RFLP) and 13 random amplification of polymorphic DNA (RAPD) markers adapted from The Bala×Azucena population includes 7 SSR, 105 RFLP and 34 AFLP markers, as well as one marker derived from oats.

#### Consensus map construction

To identify genetic map regions containing multiple overlapping QTL across populations, we computationally merged the maps of the two aforementioned mapping populations with common anchor markers using the NCGR ISYS comparative mapping tool (NCGR; http://www.ncgr.org/isys; Siepel *et al.*, 2001).

#### Rice genome sequence map

Identification of candidate genes corresponding to a QTL interval requires a contiguous genome sequence map. Although chromosome 1 and chromosome 4 were essentially completed by the International Rice Genome Sequencing Project (IRGSP) during the latter part of this project, early in the project we needed to assemble our own sequence map using available public data. This assembly work focussed on rice chromosome 1. The input data for map assembly included the list of clone names, GenBank accession numbers and the public rice physical map (Chen et al., 2002). We assembled the bacterial artificial chromosome (BAC) clone sequences into a pseudo-molecule using a suite of Perl scripts graciously provided by Dr. David Beare of the Sanger Centre, United Kingdom.

## Anchoring sequence-based genetic markers to the sequence map

We used a local implementation of the "electronic polymerase chain reaction" approach ("ePCR"; Schuler, 1997) using Perl regular expression alignment of primer sequences with orientation and threshold distance constraints to identify putative PCR amplicons in target sequences. The primers for the analysis were obtained retrieved from either the Gramene database (Ware *et al.*, 2002; www.gramene. org) or the Japan Rice Genome Project (RGP) database (Sasaki, 2001). Markers were correlated with the physical map by ePCR, run against the rice genome BAC sequences, retrieval from the Gramene database and by BLAST (Altschul *et al.*, 1990) alignment searches of the rice BAC sequences.

#### Database storage and visualization of maps

The resulting genetic (QTL), physical and sequence maps compiled for the study were stored in the International Rice Information System (IRIS; www.iris.irri.org; Bruskiewich *et al.*, 2003) and visualized using a comparative mapping tool developed at the National Center for Genome Resources. To facilitate identification of candidate genes, we also have developed QTL2Gene (http://ibi.zju.edu.cn/qtl2gene/qtl2gene.htm) as a flexible database using the public rice genome sequence and RFLP and SSR markers. The database provides an interface for searching the genes underlying one QTL using two

flanking markers' name. Other information related with this paper has been published on website (http://ibi.zju.edu.cn/publish/QTL2gene/drought.htm).

## Choice of candidate QTL region for detailed analysis

The choice of candidate QTL region on rice chromosome 1 was chosen based on a survey of drought QTL (Price *et al.*, 2002c; Lafitte *et al.*, 2002) and International Rice Research Institute (IRRI) fine mapping work (<sup>1</sup>Zhikang Li, personal communications). This region is flanked by RM212 and RM319, which were shown to be located on 148.7 cM (CentiMorgan) and 150.5 cM of IR64×Azucena genetic map respectively, and spans a map distance of approximately 1.8 cM across the sd1 gene locus.

#### Gene and functional annotation

We downloaded gene annotation for the RM212-RM319 candidate region from the Japan Rice Genome Project (RGP) sequence data found in Gen-Bank (www.ncbi.nlm.nih.gov) and from the rice databases of The Institute for Genomic Research (TIGR; www.tigr.org). These annotations were assessed using the *gbrowse* sequence browser from the Generic Model Organism Database project (http://www.gmod.org; Stein *et al.*, 2002; Lewis *et al.*, 2002).

#### Drought panicle cDNA library

An IR64-based drought stressed rice panicle cDNA library was constructed by researchers at the International Rice Research Institute (Arumugam *et al.*, 2005). Briefly, the normalized library was constructed from pooled mRNA obtained from the rice panicles collected from control (well watered) and water stressed plants at 2 d before heading, at heading, 50% flowering and 4 d after 50% flowering. Water stress was applied by not watering for several consecutive days. Expressed sequence tags (EST) obtained by 5' and/or 3' end sequencing of the clones were clustered and annotated using standard in silico approaches (Altschul *et al.*, 1990).

#### RESULTS

Alignment of the rice genetic map onto the physical map and gene identification

The chromosome 1 consensus map IR64×Azucena and Bala×Azucena was anchored using sequence-based markers to the assembled rice physical map. The candidate sequence interval spanning the candidate OTL flanked by RM212 to RM319 was then identified. The assembly of seven BAC clones in this interval yielded a contiguous sequence region of 855008 base pairs within which we identified 175 predicted gene structures. Fig.1 shows a synopsis of gene annotation categories. The RGP annotation included 48 (27%) genes annotated by functional homology to known genes, 23 (13%) pseudogenes, 24 (14%) ab initio predicted genes supported by an alignment match to an EST of unknown function, and 80 (46%) hypothetical genes predicted by ab initio algorithms.

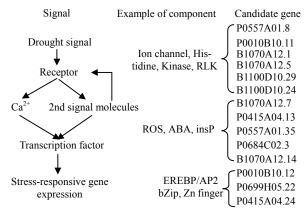


Fig.1 Possible assignment of candidate genes identified in candidate interval as components in a generic pathway for the transduction of drought stress signals in plants

## Mapping drought stressed panicle library EST into the region

To collect further supporting evidence for the function of the predicted genes in the region, EST sequences obtained from the IRRI drought stressed panicle library were aligned by BLAST (Altschul *et al.*, 1990) against candidate gene sequences. This analysis revealed that 30 candidate gene predictions match EST sequences in the library (Table 1).

## Candidate genes placed into drought signal pathways

In our study, we found 16 candidate genes in the region with some literatures support (Shinozaki and Yamaguchi-Shinozaki, 1996; Ingram and Bartels, 1996; Yamaguchi-Shinozaki *et al.*, 2002) potentially involved in drought stress response (Table 2).

<sup>&</sup>lt;sup>1</sup>Li, Z.K., 2002

Table 1 EST alignments to candidate genes

| Gene name   | Gene  |      |       |          | EST GenBank  |       |      | <ul><li>Annotation</li></ul>   |
|-------------|-------|------|-------|----------|--------------|-------|------|--|
|             | Start | Stop | score | identity | accession ID | Start | Stop | Amotation  |
| B1011A07.16 | 607   | 852  | 690   | 75.61    | CA762217     | 2     | 247  | Putative pectinesterase 2.1 precursor  |
| B1070A12.1  | 2131  | 2424 | 1380  | 96.6     | CA765415     | 1     | 294  | Putative serine/threonine kinase   |
| B1070A12.17 | 1305  | 973  | 1655  | 99.7     | CA760294     | 352   | 685  | Hypothetical protein   |
| P0010B10.6  | 1221  | 1024 | 990   | 100      | CA760259     | 488   | 685  | Hypothetical protein   |
| P0010B10.7  | 511   | 344  | 831   | 99.4     | CA763140     | 493   | 660  | Zinc protease  |
| P0010B10.12 | 1     | 249  | 1245  | 100      | CA762979     | 439   | 687  | Helicase-like transcription factor   |
| P0010B10.21 | 458   | 570  | 330   | 77.19    | CB096315     | 212   | 325  | Hypothetical protein   |
| P0010B10.21 | 458   | 570  | 321   | 76.32    | CB096974     | 175   | 288  | Hypothetical protein   |
| P0010B10.22 | 861   | 443  | 1663  | 88.54    | CA759417     | 267   | 685  | Secretory carrier membrane protein   |
| B1100D10.21 | 582   | 1    | 2868  | 99.31    | CA762908     | 135   | 715  | Acyl-CoA:1-acylglycerol-3-ph-osphate acyltransferase                               |
| B1100D10.25 | 223   | 38   | 513   | 76.72    | CB096516     | 86    | 271  | Hypothetical protein   |
| B1100D10.33 | 1790  | 1918 | 304   | 71.76    | CA764883     | 1     | 130  | Serine/threonine kinase receptor<br>precursor (EC 2.7.1.37)<br>(S-receptor kinase) |
| P0458E05.33 | 354   | 47   | 720   | 74.14    | CA759325     | 282   | 592  | Hypothetical protein   |
| P0482C06.17 | 483   | 227  | 638   | 74.81    | CA760143     | 176   | 429  | Hypothetical protein   |
| P0682C06.15 | 330   | 1    | 1533  | 96.06    | CA766823     | 326   | 655  | Putative prolyl endopeptidase  |
| P0682C06.16 | 360   | 2    | 1375  | 87.19    | CA763742     | 143   | 500  | Unknown protein  |
| P0699H05.4  | 1520  | 2043 | 2593  | 99.43    | CA766000     | 2     | 525  | Putative subtilase   |
| P0699H05.7  | 2136  | 1602 | 2524  | 97.01    | CB096421     | 317   | 851  | Putative subtilase   |
| P0699H05.7  | 1678  | 2136 | 2286  | 99.78    | CA766686     | 1     | 459  | Putative subtilase   |
| P0699H05.7  | 378   | 926  | 2414  | 93.88    | CA763105     | 162   | 714  | Putative subtilase   |
| P0699H05.9  | 120   | 44   | 162   | 74.07    | CA766969     | 204   | 279  | Hypothetical protein   |
| P0699H05.17 | 292   | 1    | 1436  | 99.32    | CB096487     | 1     | 291  | Hypothetical protein   |
| P0699H05.17 | 323   | 1    | 1606  | 99.69    | CA766130     | 35    | 357  | Hypothetical protein   |
| P0699H05.22 | 670   | 708  | 195   | 100      | CA767024     | 2     | 40   | Ethylene-responsive element binding factor 3                                       |
| P0699H05.22 | 708   | 194  | 1977  | 87.4     | CA759952     | 183   | 696  | Ethylene-responsive element binding factor 3                                       |

Table 2 Genes encoding proteins of known function possibly that may possibly be responsive to drought stress

| Classes                  | Gene name   | Putative functional description                        | Protein_id | Reference               |
|--------------------------|-------------|--|------------|-------------------------|
| Receptor                 | P0557A01.8  | Kinase-like  | BAB89770.1 | Fujimoto et al., 2000   |
|                          | P0010B10.11 | Kinase-like protein receptor                           | BAB63567.1 | Zhu, 2002               |
|                          | B1070A12.1  | Serine/threonine kinase                                | BAB92578.1 | Xiong and Zhu, 2001     |
| B1070A12.2<br>B1070A12.5 |             | Receptor kinase  | BAB92579.1 | Xiong et al., 2002      |
|                          |             | Receptor kinase  | BAB92581.1 | Xiong et al., 2002      |
|                          | B1100D10.29 | Diacylglycerol kinase                                  | BAB92552.1 | Arisz et al., 2003      |
| Ca <sup>2+</sup>         | B1100D10.24 | Cyclic nucleotide and calmodulin-regulated ion channel |            | Urao et al., 1994       |
| Signalling               | B1070A12.7  | 1,4-benzoquinone reductase                             | BAB92583.1 | Choi et al., 2002       |
|                          | P0415A04.13 | Peroxidase-like protein contains EST AU075654 (E1982)  |            | Kim et al., 2003        |
|                          | P0557A01.35 | Polyphenol oxidase                                     | BAB89784.1 | Zhang and Kirkham, 1994 |
|                          | P0684C02.3  | Polyphenol oxidase                                     | BAB89047.1 | Zhang and Kirkham, 1994 |
|                          | B1070A12.14 | Auxin-responsive GH3                                   | BAB92590.1 | Kovtun et al., 2000     |
| Transcript               | P0010B10.12 | Helicase-like transcription factor                     | BAB63568.1 | Zhu, 2002               |
|                          | P0699H05.22 | Ethylene-responsive element binding factor 3           |            | Fujimoto et al., 2000   |
|                          | P0415A04.24 | Nuclear transport factor 2                             | BAB90110.1 | Seki et al., 2002       |
| Vacuolar                 | P0010B10.27 | Vacuolar sorting-associated protein                    |            | Gaxiola et al., 2001    |

#### **DISCUSSION**

In this study, we anchored rice QTL maps to the rice physical map to identify drought stress tolerance candidate genes based on candidate QTL position. We chose the sd1 region of chromosome 1 for our attempt to link drought stress tolerance genotype to phenotype using bioinformatics because of strong combined genetic evidence for the existence of a large effect QTL for stress tolerance and because of the relatively complete rice genome sequences in the region. A set of candidate genes of known or inferred function were identified in this region using rice genome annotation. Published literature supports candidacy of some of these genes in drought stress response.

Fig.1 shows one generic drought signal pathway (Zhu, 2002) against which the candidate genes identified in this study could potentially be assigned. This pathway has three classes of genes: (1) signal perception, such as P0557A01.8; (2) generation of second messengers, such as B1070A12.7; and (3) gene expression. Candidate genes potentially implicated in this pathway are listed on the right hand side of the Fig.1.

Another possible pathway for drought signals is the Salt-Overly-Sensitive (SOS) pathway (Xiong *et al.*, 2002), as shown in Fig.2. Hypothetically, a myristoylated calcium-binding protein encoded by SOS3 may sense a drought stress or salt-elicited calcium signal and translate it downstream; SOS3 interacts with and activates SOS2, a serine/threonine protein kinase. SOS2 and SOS3 regulate the expression level of SOS1, a salt tolerance effecter gene encoding a plasma membrane Na<sup>+</sup>/H<sup>+</sup> antiporter. In our study, we found several genes that could act in this pathway: a vacuolar protein, a serine/threonine

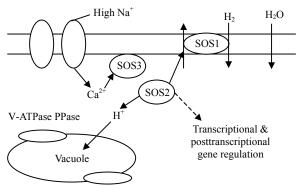


Fig.2 Regulation of ion homeostasis by the SOS pathway

kinase, three kinds of putative receptor kinase genes and a putative diacylglycerol kinase gene.

Endogenous abscisic acid (ABA) levels have been reported to increase as a result of water deficit in many physiological studies, and therefore ABA is thought to be involved in the signal transduction. ABA-dependent and ABA-independent signal pathways in the activation of stress-inducible genes under dehydration conditions have been reported (Shinozaki and Yamaguchi-Shinozaki, 1997), as shown in Fig.3. We identified an ethylene-responsive element binding factor (EREBP) gene (P0699H05.22) that could be a candidate related to this pathway. EREBP-like genes are known to be induced by a variety of abiotic stresses, including drought and chilling stresses (Gilmour *et al.*, 1998).

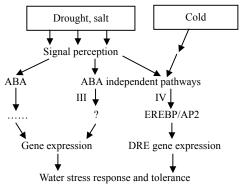


Fig.3 ABA-dependent and ABA-independent pathway

A few of the smaller gene clusters were composed of a single family of closely related genes underlying one QTL, and most of these are within 1–2 cM. Thus, we investigated this region for gene family and found that 4 rust resistance protein clusters on BAC clone (Accession number: B1100D10). These genes have above 89% identity to proteins as shown by ClastalW tool with default parameters. The rust resistance family in this region indicated that it may related to disease resistance.

One major challenge is to discover the drought candidate genes of not only those genes for which functions are already known, but also those with still unknown functions. Clearly, for such genes, fine mapping and functional genomic experimentation will be required to clarify their candidacy.

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