

1 **Genome sequence of *Bacillus subtilis* subsp. *spizizenii***  
2 **gtP20b isolated from the Indian Ocean**

3  
4  
5  
6

7 Longjiang Fan<sup>1</sup>, Shiping Bo<sup>1</sup>, Huan Chen<sup>1</sup>, Wanzhi Ye<sup>2</sup>, Katrin Kleinschmidt<sup>3</sup>, Heike I.  
8 Baumann<sup>3</sup>, Johannes F. Imhoff<sup>3</sup>, Michael Kleine<sup>4</sup> and Daguang Cai<sup>2\*</sup>

9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22

1. James D. Watson Institute of Genome Sciences & Institute of Crop Science,  
Zhejiang University, Hangzhou 310029, China
2. Molecular Phytopathology, Christian-Albrechts-Universität zu Kiel, D-24118 Kiel,  
Germany
3. Marine Microbiology, Leibniz Institute of Marine Science (IFM-GEOMAR),  
Düsternbrooker Weg 20, D-24105 Kiel, Germany
4. Planton GmbH, Am Kiel-Kanal 44, D-24106 Kiel, Germany

23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38

**Correspondence to** Prof. Dr. Daguang Cai  
Phone: +49-431-8803215  
Fax: +49-431-8801583  
E-mail: [dcai@phytomed.uni-kiel.de](mailto:dcai@phytomed.uni-kiel.de)  
Address: Department of Molecular Phytopathology,  
Christian-Albrechts-University of Kiel,  
Hermann, Rodewald Str. 9, D-24118 Kiel, Germany

39 **Key words:** Genome sequencing, *Bacillus subtilis*, marine, antimicrobial peptides, marine  
40 isolate

41 **Abstract**

42 *Bacillus subtilis* is a model organism of aerobic spore-forming Gram-positive bacteria and is  
43 of great industrial significance as the source of diverse novel functional molecules. Here we  
44 present to our knowledge the first genome sequence of a *Bacillus subtilis* strain gtP20b  
45 isolated from the marine environment. A subset of candidate genes and gene clusters were  
46 identified, which are potentially involved in production of diverse functional molecules like  
47 novel ribosomal and non-ribosomal antimicrobial peptides. The genome sequence described  
48 in this paper is due to its high strain-specificity of great importance for basic as well as  
49 applied researches on marine organisms.

50

51 **Results and discussion**

52 *Bacillus subtilis* is a member of the Gram-positive bacteria of the genus *Bacillus* and has  
53 been used as a model organism to investigate differentiation, gene/protein regulation and cell  
54 cycle events in bacteria for more than a century (1, 2). *B. subtilis* has industrial importance  
55 e.g. as a source for diverse novel functional molecules like antimicrobial peptides (2, 6).  
56 Members of the genus are ubiquitous in nature. Various strains of *B. subtilis* have been  
57 isolated from diverse habitats including seawater (4). The first *B. subtilis* genome was  
58 sequenced a decade ago (6) and updated recently (2, 9). Although draft genomes of 4 further  
59 strains were also released (8, 9), no genome of *B. subtilis* strain from a marine habitat has  
60 been decoded.

61 The *B. subtilis* subsp. *spizizenii* strain gtP20b was isolated from the sediment in 608 m  
62 water depth in the Indian Ocean and from the layer close to the ocean bottom surface. The  
63 sampling was taken through a multi corer during the cruise of the research ship Sonne at the  
64 expedition 130 in 1998 and stored at -20°C. Raw reads of the strain genome were generated  
65 by using Illumina GA (Solexa) and assembled with Velvet program (14). Based on the  
66 reference genome of *B. subtilis* strain168 (6, 7) a draft genome of gtP20b was completed. By  
67 subsequent PCR and re-sequencing 100 genome gaps were closed, but remaining four gaps  
68 and 23 unmatched short contigs (> 200 bp) with an accumulative length of 151.5 Kb, which  
69 are believed to be genome-specific and distributed in the remaining gaps.

70 The genome sequence of gtP20b comprises 4,247,908 bases with a G+C content of  
71 44.8%, and covers more than 99% of the whole genome (2, 8). It contains 4.331 open  
72 reading frames (ORFs), 77 tRNAs including one pseudogene and 30 rRNAs (3).  
73 Phylogenetic analysis revealed that gtP20b is closely clustered with *B. subtilis*168 and *B.*  
74 *natto*, but phylogenetically apart from *B. amyloliquefaciens* and *B. licheniformis* (10).  
75 Furthermore, 81.7% of the ORFs have orthologs in the strain168 (BLASTP <1e-5), but 444  
76 ORFs were not found in the released genomes of *Bacillus* genus, of these, 392 ORFs did not  
77 give hits in current public databases.

78 At least 59 genes were found to be potentially involved in secondary metabolism. They  
79 form diverse gene clusters with varied degree of synteny to other *B. subtilis* strains. A set of  
80 hits was retrieved from antimicrobial peptides (AMPs) databases (5, 11, 12, 13) including  
81 subtilisin A (*sboA*), surfactin (*sfp*), beta-lactamase precursor (*penP*) and replicative DNA  
82 helicase (*dnaC*) etc. However, they showed strong variations at both DNA- and amino acid  
83 level when compared with those of other *B. subtilis* strains, suggesting the potential of the  
84 strain gtP20b as a unique source for novel AMPs. This genome sequence is due to its high  
85 strain-specificity of great importance for both of basic and applied researches.

86

87 **Nucleotide sequence accession numbers**

88 This Whole Genome Shotgun project has been deposited at DDBJ/EMBL/GenBank  
89 under the accession AEHM00000000. The version described here is the first version under  
90 the accession number: AEHM01000000.

91

## 92 **Acknowledgements**

93 This project was supported by the Bundesministerium für Bildung und Forschung  
94 (BMBF), Germany (grant number 0315231A, B) and the Ministerium für Wissenschaft,  
95 Wirtschaft und Verkehr des Landes Schleswig-Holstein (grant number 122-08-002). Authors  
96 thanks DAAD (grant number D/08/01773, 4) and China Scholarship Council (grant number  
97 A/10/00701) for providing the scholarship reward as well as international exchange grants.  
98 Authors thank Jun Wang for his help in Solexa sequencing and Ms. Katharina Peetz for her  
99 technical support.

100

## 101 **Reference**

- 102 1. **Alcaraz, L. D., G. Moreno-Hagelsieb, L. Eguiarte, V. Souza, L. Herrera-Estrella, and**  
103 **G. Olmedo.** 2010. Understanding the evolutionary relationships and major traits of  
104 *Bacillus* through comparative. *BMC Genomics* **11**:332.
- 105 2. **Barbe, V., S. Cruveiller, F. Kunst, P. Lenoble, G. Meurice, A. Sekowska, D. Vallenet,**  
106 **T. Z. Wang, I. Moszer, C. Medigue, and A. Danchin.** 2009. From a consortium  
107 sequence to a unified sequence: the *Bacillus subtilis* 168 reference genome a decade  
108 later. *Microbiol.* **155**:1758-1775.
- 109 3. **Delcher, A.L., K. A. Bratke, E. C. Powers, and S. L. Salzberg.** 2007. Identifying  
110 bacterial genes and endosymbiont DNA with Glimmer. *Bioinformatics* **23**:673-679.
- 111 4. **Ettoumi, B., N. Raddadi, S. Borin, D. Daffonchio, A. Boudabous, and A. Cherif.** 2009.  
112 Diversity and phylogeny of culturable spore-forming *Bacilli* isolated from marine  
113 sediments. *J. Basic Microbiol.* **49**:S13-S23.
- 114 5. **Gueguen, Y., J. Garnier, L. Robert, M. P. Lefranc, I. Mougnot, J. de Lorgeril, M.**  
115 **Janech, P. S. Gross, G. W. Warr, B. Cuthbertson, M. A. Barracco, P. Bulet, A.**  
116 **Aumelas, Y. Yang, D. Bo, J. Xiang, A. Tassanakajon, D. Piquemal, and E. Bachère.**  
117 2006. PenBase, the shrimp antimicrobial peptide penaeidin database: Sequence-based  
118 classification and recommended nomenclature. *Dev. Comp. Immunol.* **30**:283-288.
- 119 6. **Kunst, F., N. Ogasawara, I. Moszer, A. M. Albertini et al.** 1997. The complete genome  
120 sequence of the Gram-positive bacterium *Bacillus subtilis*. *Nature* **390**:249-256.
- 121 7. **Kurtz, S., A. Phillippy, A. L. Delcher, M. Smoot, M. Shumway, C. Antonescu, and S.**  
122 **L. Salzberg.** 2004. Versatile and open software for comparing large genomes. *Genome*  
123 *Biol.* **5**:9.
- 124 8. **Nishito, Y., Y. Osana, T. Hachiya, K. Pependorf, A. Toyoda, A. Fujiyama, M. Itaya,**  
125 **and Y. Sakakibara.** 2010. Whole genome assembly of a natto production strain *Bacillus*  
126 *subtilis* natto from very short read data. *BMC Genomics* **11**:12.
- 127 9. **Srivatsan, A., Y. Han, J. Peng, A. K. Tehranchi, R. Gibbs, J. D. Wang, and R. Chen.**  
128 2008. High-precision, whole-genome sequencing of laboratory strains facilitates genetic  
129 studies. *PLoS Genetics*. **4(8)**: e1000139. doi:10.1371/journal.pgen.1000139.
- 130 10. **Tamura, K., J. Dudley, M. Nei, and S. Kumar.** 2007. MEGA4: Molecular evolutionary  
131 genetics analysis (MEGA) software version 4.0. *Mol. Biol. Evol.* **24**:1596-1599.

- 132 **11. Thomas, S., S. Karnik, R. S. Barai, V. K. Jayaraman, and S. Idicula-Thomas.** 2010.  
133 CAMP: a useful resource for research on antimicrobial peptides. *Nucleic Acids Res.*  
134 **38:D774-D780.**
- 135 **12. Wang, G. S., X. Li, and Z. Wang.** 2009. APD2: the updated antimicrobial peptide  
136 database and its application in peptide design. *Nucleic Acids Res.* **37:D933-D937.**
- 137 **13. Wang, C. K.L., Q. Kaas, L. Chiche, and D. J. Craik.** 2008. CyBase: a database of cyclic  
138 protein sequences and structures, with applications in protein discovery and  
139 engineering. *Nucleic Acids Res.* **36:D206-D210.**
- 140 **14. Zerbino, D. R., and E. Birney.** 2008. Velvet: Algorithms for de novo short read assembly  
141 using de Bruijn graphs. *Genome Res.* **18:821-829.**